

Exhibit 3

DECLARATION OF PAUL POLAKIS, Ph.D.

I, Paul

Polakis, Ph.D., declare and say as follows:

1. I was a graduate of the Michigan State University and received my Ph.D. from the Department of Biochemistry of the Michigan State University.

2. My scientific Curriculum Vitae is attached to and forms part of this declaration (Exhibit A).

3. I am currently employed by Genentech, Inc. where my job title is Staff Scientist. I have been leading the Tumor Antigen Project, which is a large research project aimed at identifying tumor cell markers that find use as targets for treatment of cancer in humans.

4. As part of the Tumor Antigen Project, my laboratory has been analyzing various genes in tumor cells relative to normal cells. The purpose of this project is to identify proteins that are abundantly expressed in tumor cells that are either (i) not expressed, or (ii) expressed at lower levels in normal cells. We call such differentially expressed proteins "tumor antigens". When such a tumor antigen protein is identified, we develop an antibody that recognizes and binds to that protein. Such an antibody can be used in the diagnosis of human cancer and may ultimately serve as a target for treatment in the treatment of human cancer.

5. In the course of the research conducted by Genentech's Tumor Antigen Project, we have used a variety of scientific techniques for detecting and measuring gene expression in human tumor cells relative to normal cells, including protein levels. An important example of one such technique is the widely used technique of microarray analysis which has been extremely useful for the identification of mRNA molecules that are over-expressed in one tissue or cell type relative to another. In the course of microarray analysis, we have identified approximately 1000 transcripts that are present in human tumor cells at levels that are significantly higher than in corresponding normal human cells. To date, we have generated antibodies that bind to about 30 of the tumor antigen proteins and have used these antibodies to determine the level of production of these tumor antigens in cancer cells and corresponding normal cells. We have also performed mRNA and protein expression analyses described in paragraph 4 above, where there is a strong correlation between changes in the level of mRNA and the level of protein.

6. The results of the research conducted by Genentech's Tumor Antigen Project, which has been extremely useful for the identification of mRNA molecules that are over-expressed in one tissue or cell type relative to another. In the course of microarray analysis, we have identified approximately 1000 transcripts that are present in human tumor cells at levels that are significantly higher than in corresponding normal human cells. To date, we have generated antibodies that bind to about 30 of the tumor antigen proteins and have used these antibodies to determine the level of production of these tumor antigens in cancer cells and corresponding normal cells. We have also performed mRNA and protein expression analyses described in paragraph 4 above, where there is a strong correlation between changes in the level of mRNA and the level of protein.

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experience accumulated in more than 20 years of
discussed in paragraphs 4 and 5 above and my
scientific literature, it is my considered scientific
opinion, an increased level of mRNA in a tumor cell relative
correlates to a similar increase in abundance of the
protein relative to the normal cell. In fact, it remains a
fact of biology that increased mRNA levels are predictive of
levels of the encoded protein. While there have been
cases in which such a correlation does not exist, it is my
opinion that exceptions to the commonly understood general rule
are predictive of corresponding increased levels of the

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All statements made herein of my own knowledge are
made on information or belief are believed to be true,
statements were made with the knowledge that willful false
statements are punishable by fine or imprisonment, or both,
under 18 of the United States Code and that such willful
statements are invalid for the purpose of the application or any patent issued

Dated: 5/

By: Paul Polakis

Paul Polakis, Ph.D.

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CURRICULUM VITAE

PAUL G. POLAKIS

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San Francisco, CA 94080

EDUCATION:

Ph.D., Biochemistry,
Michigan State University

Biochemistry,

B.S., Biology, College

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PROFESSIONAL

2002-present

Staff Scientist, Genentech, Inc
S. San Francisco, CA

1999- 2002

Senior Scientist, Genentech, Inc.,
S. San Francisco, CA

1997 -1999

Research Director
Onyx Pharmaceuticals, Richmond, CA

1992- 1996

Senior Scientist, Project Leader, Onyx
Pharmaceuticals, Richmond, CA

1991-1992

Senior Scientist, Chiron Corporation,
Emeryville, CA.

1989-1991

Scientist, Cetus Corporation, Emeryville CA.

1987-1989

Postdoctoral Research Associate, Genentech,
S. San Francisco, CA.

1985-1987

Postdoctoral Research Associate, Department
of Medicine, Duke University Medical Center,
Durham, NC

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1984-1985

Assistant Professor, Department of Chemistry,
Oberlin College, Oberlin, Ohio

1980-1984

Graduate Research Assistant, Department of
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PUBLICATIONS:

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Wu TD, Zhou Y, Watanabe C, Luoh SM, Polakis P,
quantitative gene expression profiling in normal and

Genome-wide Study of Gene Copy Numbers, Protein Levels in Pairs of Non-invasive and Invasive Human Transitional Cell Carcinoma

Torben F. Hansen¹, Søren Thomsen¹, and Julio E. Celis²

Frederic M. Waldman³, Hans Wolf⁴

Gain and loss of chromosomes of bladder cancer is well a general. The consequences transcription and translation partly because of technical difficulties attempted to address this question in non-invasive and invasive human bladder tumors. of technology included oligonucleotide hybridization, high density oligonucleotide arrays, and two-dimensional electrophoresis. It is concluded that there is a dose-dependent effect of gene copy number on protein levels. In a comparative genomic hybridization (CGH) study of 23 cases, we found that the loss of chromosome 9 was associated with a decrease in the level of DNA-dependent protein kinase (p34^{cdc2}). This loss showed that the effect of gene copy number on protein levels is not only present in the non-invasive but also in the invasive disease. The correlation between gene copy number and protein levels of the app. 2000

phenomenon at both the transcription and translation levels. High throughput array studies of the breast cancer cell line BT474 has suggested that there is a correlation between DNA copy numbers and gene expression in highly amplified areas (2), and studies of individual genes in solid tumors have revealed a good correlation between gene dose and mRNA or protein levels in the case of *c-erb-B2*, *cyclin D1*, *ras*, and *N-myc* (3–5). However, a high cyclin D1 protein expression has been observed without simultaneous amplification (4), and a low level of *c-myc* copy number increase was observed without concomitant *c-myc* protein overexpression (6).

In human bladder tumors, karyotyping, fluorescent *in situ* hybridization, and comparative genomic hybridization (CGH)¹ have revealed chromosomal aberrations that seem to be characteristic of certain stages of disease progression. In the case of non-invasive pTa transitional cell carcinomas (TCCs), this includes loss of chromosome 9 or parts of it, as well as loss of Y in males. In minimally invasive pT1 TCCs, the following alterations have been reported: 2q-, 11p-, 1q+, 11q13+, 17q+, and 20q+ (7–12). It has been suggested that these regions harbor tumor suppressor genes and oncogenes; however, the large chromosomal areas involved often contain many genes, making meaningful predictions of the functional consequences of losses and gains very difficult.

In this investigation we have combined genome-wide technology for detecting genomic gains and losses (CGH) with gene expression profiling techniques (microarrays and proteomics) to determine the effect of gene copy number on transcript and protein levels in pairs of non-invasive and invasive human bladder TCCs.

EXPERIMENTAL PROCEDURES

Material—Bladder tumor biopsies were sampled after informed consent was obtained and after removal of tissue for routine pathology examination. By light microscopy tumors 335 and 532 were staged by an experienced pathologist as pTa (superficial papillary).

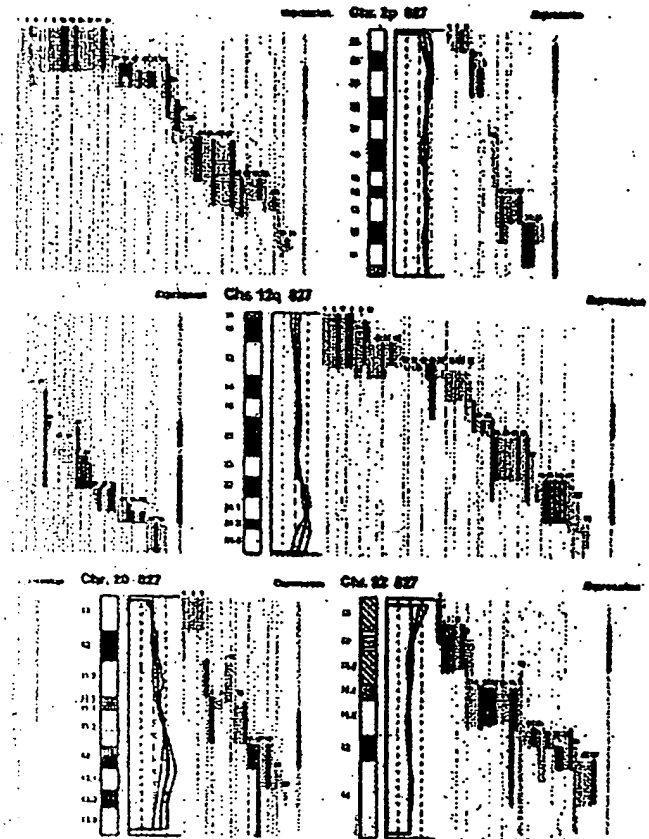
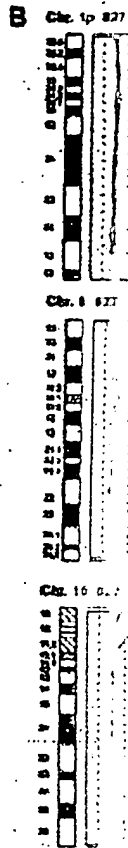
¹ The abbreviations used are: CGH, comparative genomic hybridization; TCC, transitional cell carcinoma; LOH, loss of heterozygosity; PA-FABP, psoriasis-associated fatty acid-binding protein; 2D, two-dimensional.

Aneuploidy (1), but little

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for 30 min at 70 °C. The probe arrays were then scanned by a microarray scanner (GeneChip Reader) from the Affymetrix Corporation.

Microarray analysis. The data were analyzed using the software described previously (www.noble.org). The data were obtained from the Affymetrix GeneChip Reader. The data were analyzed using the software described previously (www.noble.org). The data were obtained from the Affymetrix GeneChip Reader.

Protein analysis. The protein samples were homogenized in RNeasy lysis buffer. Samples were then subjected to SDS-PAGE. Samples were then subjected to SDS-PAGE. Samples were then subjected to SDS-PAGE.

CGH—Hy. The CGH analysis was performed as previously described.

labeled reference DNA (200 ng), and human Col-1 DNA (20 µg) were denatured at 37 °C for 5 min and applied to denatured normal metaphase slides. Hybridization was at 37 °C for 2 days. After washing, the slides were counterstained with 0.15 µg/ml 4,6-diamidino-2-phenylindole in an anti-fade solution. A second hybridization was performed for all tumor samples using fluorescein-labeled reference DNA and Texas Red-labeled tumor DNA (inverse labeling) to confirm the aberrations detected during the initial hybridization. Each CGH experiment also included a normal control hybridization using fluorescein- and Texas Red-labeled normal DNA. Digital image analysis was used to identify chromosomal regions with abnormal fluorescence ratios, indicating regions of DNA gains and losses. The average green:red fluorescence intensity ratio profiles were calculated using four images of each chromosome (eight chromosomes total) with normalization of the green:red fluorescence intensity ratio for the entire metaphase and background correction. Chromosome identification was performed based on 4,6-diamidino-2-phenylindole banding patterns. Only images showing uniform high intensity fluorescence with minimal background staining were analyzed. All centromeres, p arms of acrocentric chromosomes, and heterochromatic regions were excluded from the analysis.

RESULTS

Comparative Genomic Hybridization—The CGH analysis identified a number of chromosomal gains and losses in the

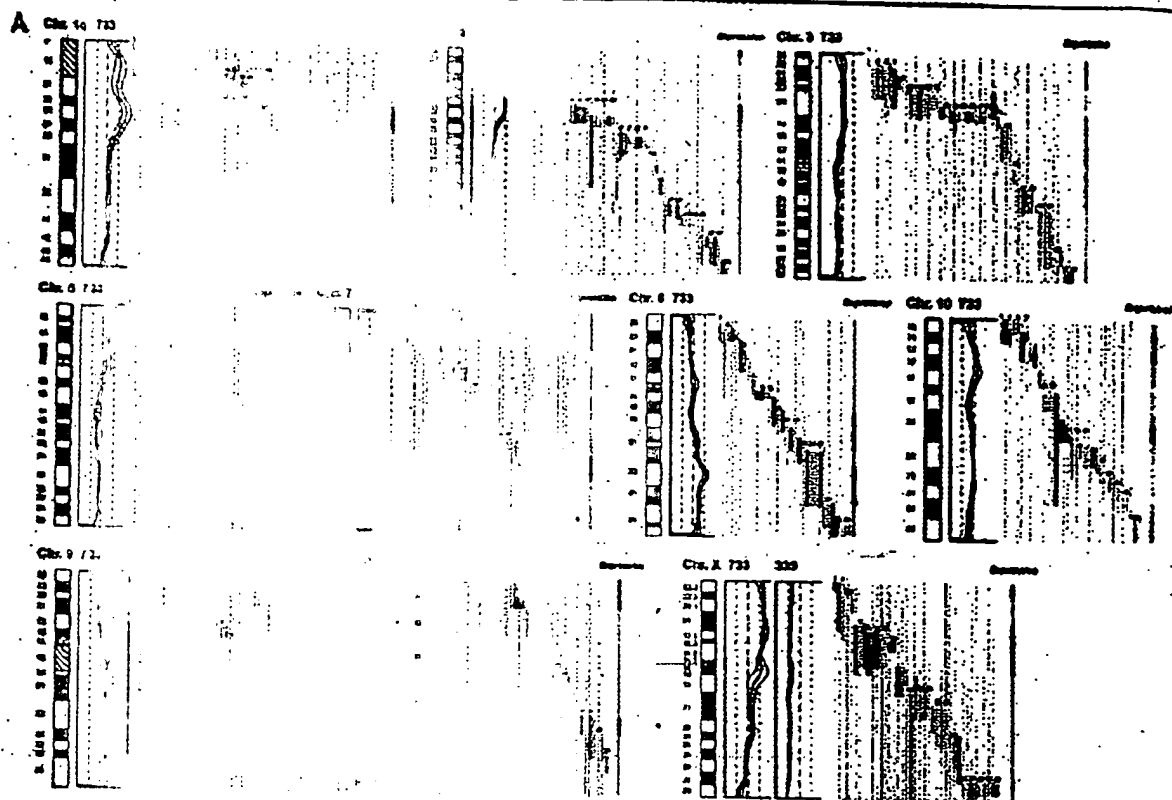


FIG. 1. Comparison of gene expression and DNA content in invasive and non-invasive tumors. A, expression of mRNA in invasive tumor 733 compared with the non-invasive counterpart (B). The left column shows the CGH profiles of the chromosomes. The colored bars represent the ratio of DNA content in the invasive tumor compared with the non-invasive counterpart. The bars indicate the purported location of the gene. The bar to the far right, entitled Expression, shows the resulting change in expression of the genes. The bar to the far left, entitled Chromosome, shows the chromosome. The bar to the far right, entitled Expression, shows the resulting change in expression of the genes. The bar to the far left, entitled Chromosome, shows the chromosome.

grade I and II (invasive and non-invasive) staged as pT1a and pT1b. mRNA levels were determined by Northern blot analysis. The first and second round of Northern blot analysis were performed using the same Northern blotting protocol. The first and second round of Northern blot analysis were performed using the same Northern blotting protocol.

From left to right are chromosome (Chr.), CGH profiles, gene location and gene expression. A, expression of mRNA in invasive tumor 733 compared with the non-invasive counterpart (B). The left column shows the CGH profiles of the chromosomes. The colored bars represent the ratio of DNA content in the invasive tumor compared with the non-invasive counterpart. The bars indicate the purported location of the gene. The bar to the far right, entitled Expression, shows the resulting change in expression of the genes. The bar to the far left, entitled Chromosome, shows the chromosome. The bar to the far right, entitled Expression, shows the resulting change in expression of the genes. The bar to the far left, entitled Chromosome, shows the chromosome.

UTP (Enzo) was used, together with unlabeled NTPs in the reaction. Following the *in vitro* transcription reaction, the unincorporated nucleotides were removed using RNeasy columns (Qiagen).

Array Hybridization and Scanning.—Array hybridization and scanning was modified from a previous method (13). 10 μ g of cRNA was fragmented at 94 °C for 35 min in buffer containing 40 mM Tris acetate, pH 8.1, 100 mM KOAc, 30 mM MgOAc. Prior to hybridization, the fragmented cRNA in a 6X SSPE-T hybridization buffer (1 M NaCl, 10 mM Tris, pH 7.6, 0.005% Triton), was heated to 95 °C for 5 min, subsequently cooled to 40 °C, and loaded onto the Affymetrix probe array cartridge. The probe array was then incubated for 18 h at 40 °C at constant rotation (60 rpm). The probe array was exposed to 10 washes in 6X SSPE-T at 25 °C followed by 4 washes in 0.5X SSPE-T at 50 °C. The biotinylated cRNA was stained with a streptavidin-phycoerythrin conjugate, 10 μ g/ml (Molecular Probes) in 6X SSPE-T.

Gene Copy		Correlation		Table 1	
Top, CGH independent		Independent		CGH alterations used by CGH and by expression monitoring	
CGH alterations		73%		what expression ratio was found; bottom, altered expression used as what expression ratio was found.	
13 Gain		10 Loss		CGH alterations	
10 Loss		10 Loss		Tumor 827 vs. 532	
10 Loss		10 Loss		Expression change clusters	
10 Loss		10 Loss		Concordance	
10 Loss		10 Loss		10 Gain	
10 Loss		10 Loss		8 Up-regulation	
10 Loss		10 Loss		0 Down-regulation	
10 Loss		10 Loss		2 No change	
10 Loss		10 Loss		3 Up-regulation	
10 Loss		10 Loss		2 Down-regulation	
10 Loss		10 Loss		7 No change	
10 Loss		10 Loss		17 Up-regulation	
10 Loss		10 Loss		5 Loss	
10 Loss		10 Loss		2 No change	
10 Loss		10 Loss		0 Gain	
10 Loss		10 Loss		3 Loss	
10 Loss		10 Loss		8 No change	
10 Loss		10 Loss		1 Gain	
10 Loss		10 Loss		3 Loss	
10 Loss		10 Loss		17 No change	
10 Loss		10 Loss		17 Up-regulation	
10 Loss		10 Loss		5 Loss	
10 Loss		10 Loss		2 No change	
10 Loss		10 Loss		0 Gain	
10 Loss		10 Loss		3 Loss	
10 Loss		10 Loss		8 No change	
10 Loss		10 Loss		1 Gain	
10 Loss		10 Loss		3 Loss	
10 Loss		10 Loss		17 No change	

two invasive the two non 532) show and Y-, res. (1q22-24+, 9q34+, 11, 13+, 14, 15, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 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change in ratio between invasive tumors 827 (▲) and 733 (◆) and their non-invasive counterparts taken from the *Expressioni* line to the right in Fig. 1, which depicts the resulting fold change of the mRNAs from a given region have to be either up- or down-regulated in the invasive arms in which the CGH ratio plus or minus one standard deviation was outside the

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In both TCC 733 and TCC 827, the telomeric end of chromosome 11p showed a normal ratio in the CGH analysis; however, clusters of five and three genes, respectively, lost their expression. Two microsatellites (D11S1760, D11S922) positioned close to MUC2, IGF2, and cathepsin D indicated LOH as the most likely mechanism behind the loss of expression (data not shown).

A reduced expression of mRNA observed in TCC 733 at chromosomes 3q24, 11p11, 12p12.2, 12q21.1, and 16q24 and in TCC 827 at chromosome 11p15.5, 12p11, 15q11.2, and 18q12 was also examined for chromosomal losses using microsatellites positioned as close as possible to the gene loci

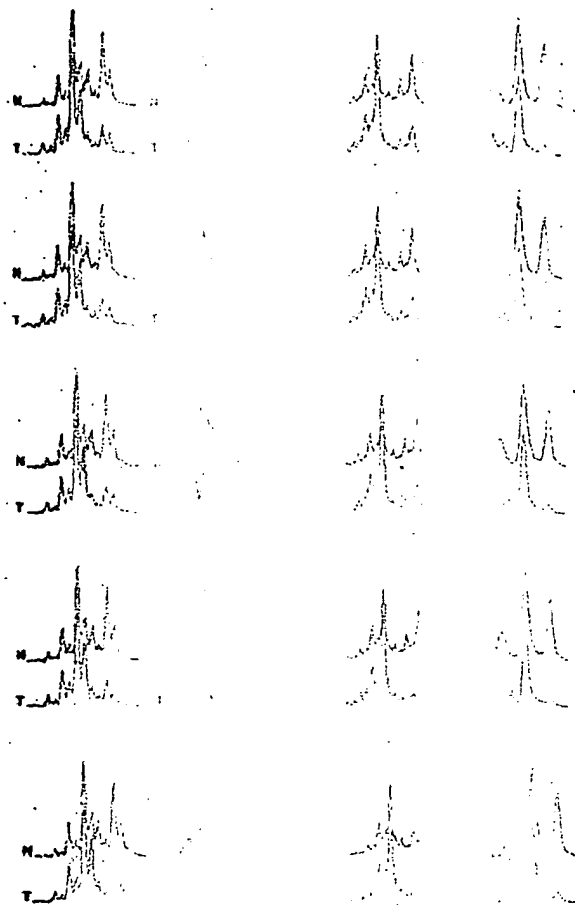


Fig. 3. Northern blot analysis of mRNA levels for TCCs 733 and 532. (a) by DISC analysis, number 38 (TCC 733) and number 41 (TCC 532) to general β -actin showing localization close to mitochondrial DNA from normal cells. (b) electrophoresis partially lost

showing repositioned transcripts may be observed. Relation 2D-PAGE. Blue and/or using fresh proteins

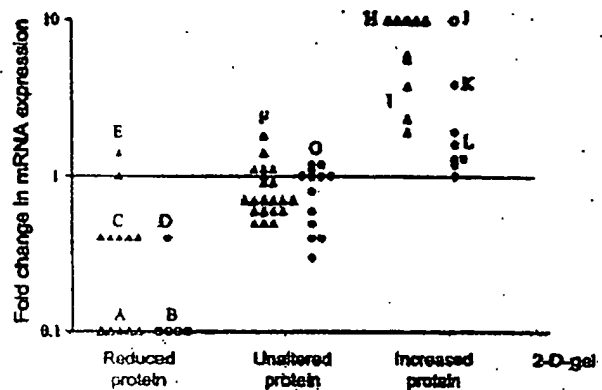


Fig. 4. Correlation between protein levels as judged by 2D-PAGE and transcript ratio. For comparison proteins were divided in three groups, unaltered in level or up- or down-regulated (horizontal axis). The mRNA ratio as determined by oligonucleotide arrays was plotted for each gene (vertical axis). Δ , mRNAs that were scored as present in both tumors used for the ratio calculation; Δ , mRNAs that were scored as absent in the invasive tumors (along horizontal axis) or as absent in non-invasive reference (top of figure). Two different scalings were used to exclude scaling as a confounder, TCCs 827 and 532 (Δ) were scaled with background suppression, and TCCs 733 and 335 (\bullet) were scaled without suppression. Both comparisons showed highly significant ($p < 0.005$) differences in mRNA ratios between the groups. Proteins shown were as follows: Group A (from left), phosphoglucosylase-1, glutathione transferase class μ number 4, fatty acid-binding protein homologue, cytochrome 15, and cytochrome 13; B (from left), fatty acid-binding protein homologue, 28-kDa heat shock protein, cytochrome 13, and calyculin; C (from left), α -enolase, hnRNP B1, 28-kDa heat shock protein, 14-3-3 ϵ , and pre-mRNA splicing factor; D, mesothelial keratin K7 (type II); E (from top), glutathione S-transferase- π and mesothelial keratin K7 (type II); F (from top and left), adenyl cyclase-associated protein, E-cadherin, keratin 19, calgizarin, phosphoglycerate mutase, annexin IV, cytochrome γ -actin, hnRNP A1, integral membrane protein calnexin (IP90), hnRNP H, brain-type clathrin light chain-a, hnRNP F, 70-kDa heat shock protein, heterogeneous nuclear ribonucleoprotein A/B, translationally controlled tumor protein, liver glyceraldehyde-3-phosphate dehydrogenase, keratin 8, aldehyde reductase, and Na,K-ATPase β -1 subunit; G, (from top and left), TCP20, calgizarin, 70-kDa heat shock protein, calnexin, hnRNP H, cytochrome 15, ATP synthase, keratin 19, triosephosphate isomerase, hnRNP F, liver glyceraldehyde-3-phosphate dehydrogenase, glutathione S-transferase- π , and keratin 8; H (from left), plasma gelsolin, autoantigen calreticulin, thioredoxin, and NAD $^{+}$ -dependent 15 hydroxyprostaglandin dehydrogenase; I (from top), prollyl 4-hydroxylase β -subunit, cytochrome 13, cytochrome 17, prothibition, and fructose 1,6-bisphosphatase; J, annexin II; K, annexin IV; L (from top and left), 90-kDa heat shock protein, prollyl 4-hydroxylase β -subunit, α -enolase, GRP 78, cyclophilin, and cofilin.

gradient, and having a known chromosomal location, were selected for analysis in the TCC pair 827/532. Proteins were identified by a combination of methods (see "Experimental Procedures"). In general there was a highly significant correlation ($p < 0.005$) between mRNA and protein alterations (Fig. 4). Only one gene showed disagreement between transcript alteration and protein alteration. Except for a group of cyto-

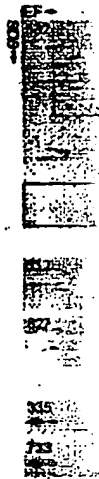


Fig. 5. C and non-invasive (left) and invasive (right) TCCs on the identical array. Clearly, cytokeratin 17 (red) and cytokeratin 20 (red) are down-regulated in the invasive TCCs and is consistent with the mRNA data. PA-FABP (red) is up-regulated in the invasive TCCs and is consistent with the mRNA data.

keratins encoded by genes on chromosome 17 (Fig. 5) the analyzed proteins did not belong to a particular family. 28 well focused proteins whose genes had a known chromosomal location were detected in TCCs 733 and 335, and of these 19 correlated ($p < 0.005$) with the mRNA changes detected using the arrays (Fig. 4). For example, PA-FABP was highly expressed in the non-invasive TCC 335 but lost in the invasive counterpart (TCC 733; see Fig. 5). The smaller number of proteins detected in both 733 and 335 was because of the smaller size of the biopsies that were available.

11 chromosomal regions where CGH showed aberrations that corresponded to the changes in transcript levels also showed corresponding changes in the protein level (Table II). These regions included genes that encode proteins that are found to be frequently altered in bladder cancer, namely cytokeratins 17 and 20, annexins II and IV, and the fatty acid-binding proteins PA-FABP and FABP1. Four of these proteins were encoded by genes in chromosome 17q, a frequently amplified chromosomal area in invasive bladder cancers.

DISCUSSION

Most human cancers have abnormal DNA content, having lost some chromosomal parts and gained others. The present study provides some evidence as to the effect of these gains and losses on gene expression in two pairs of non-invasive and invasive TCCs using high throughput expression arrays and proteomics, in combination with CGH. In general, the results showed that there is a clear individual regulation of the mRNA expression of single genes, which in some cases was superimposed by a DNA copy number effect. In most cases, genes located in chromosomal areas with gains often exhibited increased mRNA expression, whereas areas showing losses showed either no change or a reduced mRNA expression. The latter might be because of the fact that losses most often are restricted to loss of one allele, and the cut-off point for detection of expression alterations was a 2-fold change, thus being at the border of detection. In several cases, how-

TABLE II
Genes with both mRNA and gene dose changes

TCC	CGH alteration	Transcript alteration ^a	Protein alteration
	Gain	Abs to Pres ^b	Increase
	Gain	3.9-Fold up	Increase
	Gain	3.8-Fold up	Increase
	Gain	5.6-Fold up	Increase
	Loss	10-Fold down	Decrease
	Gain	2.3-Fold up	Increase
	Gain	Abs to Pres	Increase
	Loss	2.5-Fold up	Decrease
	Gain	3.7-12.5-Fold up ^b	Increase
	Gain	5.7-11.8-Fold up	Increase
	Loss	2.5-Fold down	Decrease

^a Abs, absent; Pres, present.

^b In TCCs 827 and 733 these are shown as 827/733.

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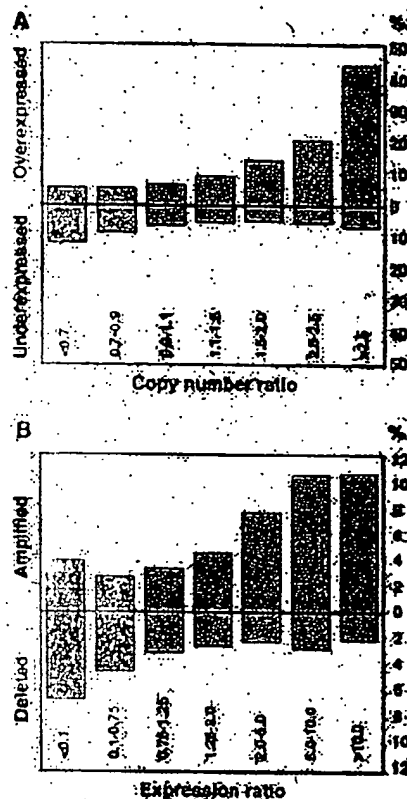
Genetic changes
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of breast cancer a

Gene expression facilitated classification; some of which (1-6). Despite this, mechanisms underlying the maintenance, clonal identification of the tumor. Accumulation of mutations in the evolution of cancer. The role of genetic changes are actively investigated. Some encoded proteins are demonstrated by immunofluorescently defined oncogenes, such as other solid tumor.

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U.S. and Biomedicum Bloch Center, Helsinki University Hospital, Biomedicum



copy number on global gene expression levels. 4, percentage of lncRNAs and genes (Y axis) according to copy number (X axis). 5, upregulated and underexpression were >2.184 (global upper 7% of os) and <0.4675 (global lower 7% of the expression ratios). 6, percentage of lncRNAs and genes according to expression ratios. Threshold values for up and down were >1.5 and <0.7.

recurrent regions of DNA amplification have been mapped in colorectal cancer by CGH³ (9, 10). However, these amplicons are often poorly defined, and their impact on gene expression remains

hypothesis that genome-wide identification of those gene changes that are attributable to underlying gene copy alterations would highlight transcripts that are actively involved in the causation or maintenance of the malignant phenotype. By such transcripts, we applied a combination of cDNA and microarrays to: (a) determine the global impact that gene copy alterations plays in breast cancer development and progression; (b) identify and characterize those genes whose mRNA expression

lative and c: CGH, comparative genomic hybridization; FISH, fluorescence in situ hybridization; RT-PCR, reverse transcription-PCR.



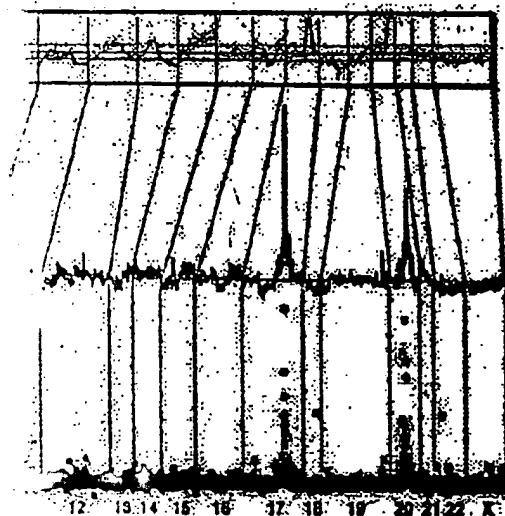
Fig. 2. Genotype
(line) across the entire
and green line, a ratio
of the cDNA clones at
copy number ratio of 1:
the next 5% of the ex,
(underexpressed genes);
indicated with a dash:

sion is most significant for the responding genotype.

MATERIALS. A

Breast Cancer Co.
474, HCC1428, Hs57-
SKBR-3, T-47D, U-
American Type Co.
recommended cu-
using standard 31

Copy Number
preparation and performed as described previously [15]. Briefly, 20 µg of digested human WBCs (Cytogenetics, Inc., Rockville, MD) was hybridized with the Bioprime Labeling kit (Amersham Pharmacia) and the posthybridization analysis, a standard La Jolla, CA) was labeled with Cy3-labeled cDNAs were microarray analysis (Alto, CA) was used locations using the average intensity of the copy number distribution of the array. Low quality intensity <100 reference intensities.



mosomal CGH analysis of MCF-7. The copy number ratio profile (Otsu). The black horizontal line indicates a ratio of 1.0; red line, a ratio of 0.8; array. The copy number ratios were plotted as a function of the position moving median of 10 adjacent clones is shown. Red horizontal line, the ratio ratios. The bright red dots indicate the upper 2%, and dark red dots, the lowest 2%, and dark green dots, the next 5% of the expression ratios. Sub is and shown at the bottom of the figure, and chromosome boundaries are

d from the analysis and were treated as missing values. The fluorescence ratios were used to define cutpoints for increased/decreased copy number. Genes with CGH ratio >1.43 (representing the upper 1% of ratios across all experiments) were considered to be amplified, and ratio <0.73 (representing the lower 5%) were considered to be

Analysis of CGH and cDNA Microarray Data. To evaluate the effect of copy number alterations on gene expression, we applied the statistical approach. CGH and cDNA calibrated intensity ratios were used and normalized using median centering of the values in each chromosome. Furthermore, cDNA ratios for each gene across all 14 cell lines were used. For each gene, the CGH data were represented by a vector with 1 for amplification (ratio, >1.43) and 0 for no amplification. This was correlated with gene expression using the signal-to-noise ratio. We calculated a weight, w_g , for each gene as follows:

$$w_2 = \frac{m_{y1} - m_{y0}}{\sigma_{y1} + \sigma_{y0}}$$

μ_1 and μ_0 denote the means and SDs for the expression of a gene in nonplasmidified cell lines, respectively. To assess the significance of each weight, we performed 10,000 random permutation tests. The probability that a gene had a larger or equal permutation value than the original weight was denoted by α . An α value indicates a strong association between gene expression and

Localization of cDNA Clones and Amplicon Mapping. Each clone in the microarray was assigned to a Unigene cluster using the 141.⁶ A database of genomic sequence alignment information generated was created from the August 2001 freeze of the Unigene Set.⁷ The chromosome and coordinates of each cDNA clone were then retrieved by relating these data to the Unigene database. Clones were then classified as CGH copy number ratio >2.0 in at least two cell lines or as CGH ratio >2.0 in at least three cell lines. The amplicon start and end positions were

url: http://research.nhgri.nih.gov/microarray/downloadable_cdna.html
url: <http://www.genome.ucsf.edu>

Table 1. Summary

Location
1p13
1q21
1q22
3p14
7p12.1-7p11.2
7q31
7q32
8q21.11-8q21.1
8q21.3
8q23.3-8q24.1
8q24.22
9p13
13q22-q31
16q22
17q11
17q12-q21.2
17q21.32-q21.1
17q22-q23.3
17q23.3-q24.3
19q13
20q11.22
20q13.12
20q13.12-q13.1
20q13.2-q13.32

extended to include
plasmid size data.

FISH. Dual-color
described (17). Slides
labeled with Spectra
Orange-labeled probes
labeled chromosome
reference. A tissue
died primary breast
(18). The use of the
University of British
increase in the
centromere signal
amplified. Survival
and the log-rank

RT-PCR. Total
GAPDH, Reverse
Access RT-PCR
as a template. Forward
and 5'-GCGTCAC

RESULTS

Global Expression
arrayed cDNA
and gene copy
lines. The results
on gene expression
transcripts (CCH)
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genes with no
of the transcripts
showed increased
increases and
less dramatic

Identification
locations obtained
number changes
ment Fig. A).
was 267 kb.
breast cancer
1). Several

validated, with 1q21, 17q12-q21.2, 17q22-q23, 20q13.1,
2 regions being most commonly amplified. Furthermore,
ries of these amplicons were precisely delineated. In ad-
del amplicons were identified at 9p13 (38.65-39.25 Mb),
3 (52.47-55.80 Mb).

Identification of Putative Amplification Target Genes.
/CGH microarray technique enables the direct correla-
y number and expression data on a gene-by-gene basis
the genome. We directly annotated high-resolution
with gene expression data using color coding. Fig. 2C
most of the amplified genes in the MCF-7 breast cancer
1p13, 17q22-q23; and 20q13 were highly overex-
view of chromosome 7 in the MDA-468 cell line
EGFR as the most highly overexpressed and amplified
1-p12 (Fig. 3A). In BT-474, the two known amplicons
nd 17q22-q23 contained numerous highly overex-
es (Fig. 3B). In addition, several genes, including the
genes *HOXB2* and *HOXB7*, were highly amplified in a
un described independent amplicon at 17q21.3. *HOXB7*
stically amplified (as validated by FISH, Fig. 3B, inset)
verexpressed (as verified by RT-PCR, data not shown)
UACC812, and ZR-75-30 cells. Furthermore, this novel

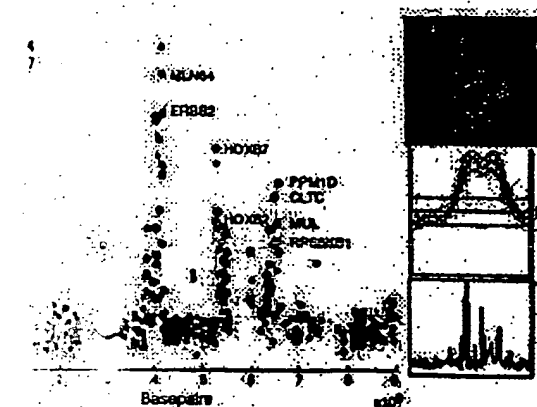
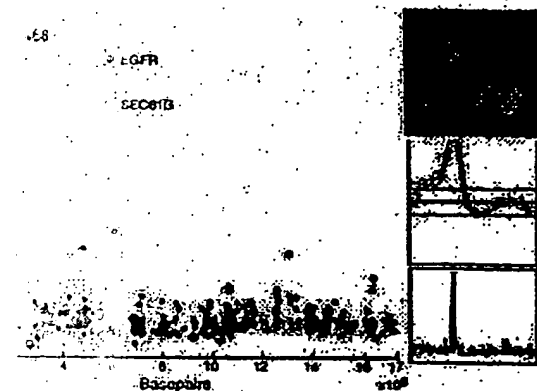
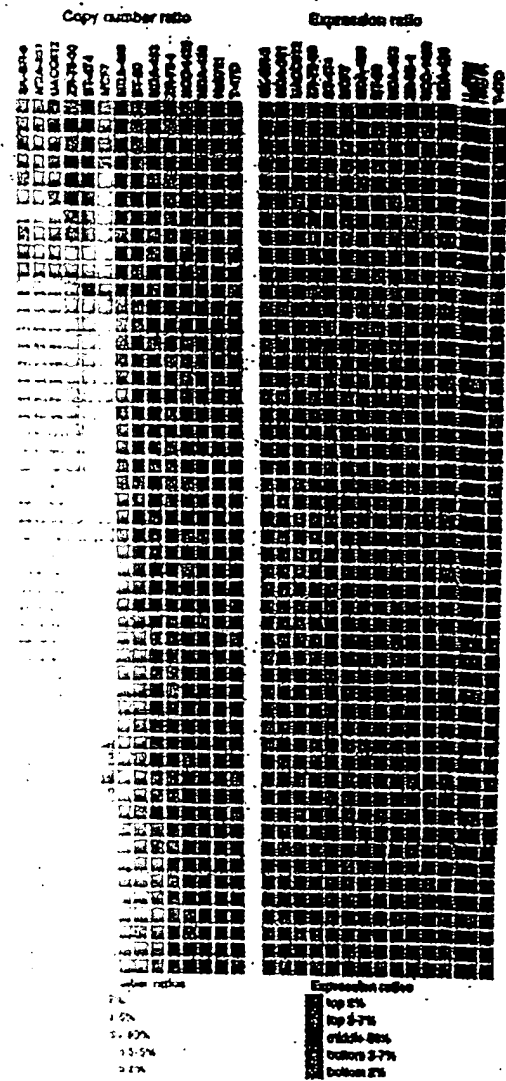


Figure 3. Validation of gene expression data on CGH microarray profiles. A, genes in the MDA-468 cell line are highly expressed (red dots) and include *EGFR*. B, several genes in the 17q12, 17q21.3, and 17q23 amplicons in the BT-474 cell line are highly overexpressed (red) and include the *HOXB7* gene. Labels and color coding are as indicated for Fig. 2C. Insets show FISH profiles for the corresponding chromosomes and validation of the number of interphase FISH using *EGFR* (red) and chromosome 7 centromere (green) in MDA-468 (A) and *HOXB7*-specific probe (red) and chromosome 7 centromere (green) in BT-474 cells (B).

Fig. 4. List of significant correlations between gene copy number and expression ratio. Genes are indicated according to their position on the right. Expression ratio put key to the color scale graph. Gray squares list of 270 genes is



amplification with breast cancers with poor prognosis. Statistical analysis of the expressed genes showed that 179 had associations (84%) are implied, and transcription, and translation that could not

current gene and chromosome copy number and progression of solid tumors has been publications applying CGH⁹ (9, 10), as well as other molecular cytogenetic, cytogenetic, and genetic studies. The effects of these somatic genetic alterations have remained largely unknown, we explored gene expression changes occurring in (15, 19-21). Here, we applied genome-wide analysis to identify transcripts whose expression is altered by underlying gene copy number alterations

copy number on gene expression patterns was dramatic effects seen in the case of high-

* Internet address

In summary, to the analysis of 12,000 transcripts once every prominent expression of amplicons in genes, the gene amplification 17q21.3 in *HOXB7* gene

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for direct role of the transcriptional

Cell^{1,2}, Stefanie S. Jeffrey^{1,2}, Per E. Lonning^{2,3},
and D. Brown^{1,2,3}

¹Howard Hughes Medical Institute, Stanford
Hospital, Montebrillo, N-0310 Oslo, Norway;
²Department of Genetics and Ueberberg

we have identified a significant impact of wide-
copy number alteration on the transcriptional
tumor.

Methods

Cell Lines. Primary breast tumors were predominantly
intermediate-grade, infiltrating ductal carcinoma,
with 50% being lymph node positive. The
cells within specimens averaged at least 50%.
Individual tumors have been published (8, 9), and
in Table 1, which is published as supporting
the PNAS web site, www.pnas.org. Breast cancer
obtained from the American Type Culture
Genomic DNA was isolated either using Qiagen
columns, or by phenol/chloroform extraction
and precipitation.

Microarray Hybridizations. Genomic DNA label-
ings were performed essentially as described
(7), with slight modifications. Two micrograms
in a total volume of 50 microliters and the
agents were adjusted accordingly. "Test" DNA
(cell lines) was fluorescently labeled (Cy5) and
human cDNA microarray containing 6,691
human genes (i.e., UniGene clusters). The
labeled with Cy3 for each hybridization was nor-
malized DNA from a single donor. The fabrication
of arrays and the labeling and hybridization of
have been described (8).

Map Positions. Hybridized arrays were scanned
per (Axon Instruments, Foster City, CA), and
(test/reference) calculated using SCANALYZE
at <http://rana.lbi.gov>. Fluorescence ratios
for each array by setting the average log
for all array elements equal to 0. Measure-
ment intensities more than 20% above back-
ground were reliable. DNA copy number profiles
significantly from background ratios measured in
DNA control hybridizations were interpreted as
DNA copy number alteration (see *Estimating
and Fluorescence Ratios* in the supporting
indicated, DNA copy number profiles are
average (symmetric 5-nearest neighbors),
arrayed human cDNAs were assigned by

Genomic hybridization.

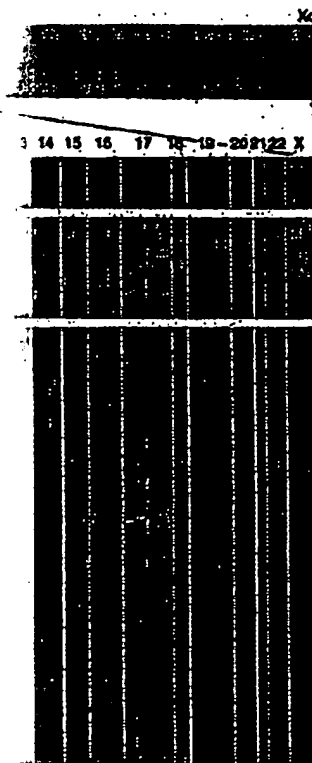
should be addressed at: Department of Pathology, Stanford
Hospital, CCSF Building, Room 3245A, 269 Campus Drive, Stanford,
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Fig. 1. G
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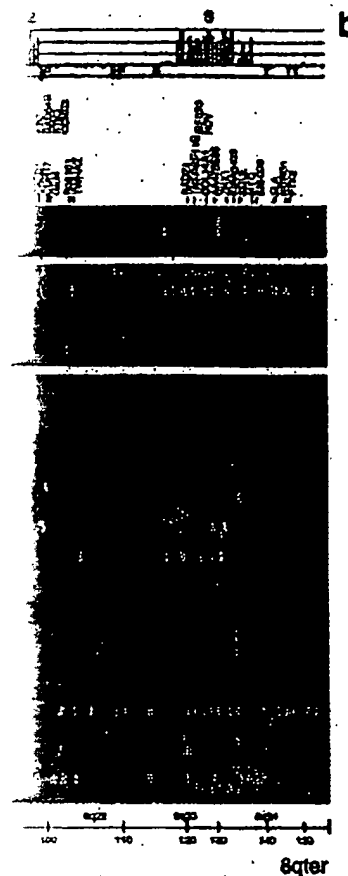
profiles are illustrated for cell lines containing different
different cell line or tumor, and each column represents
from 1pter through Xqter. Moving average (symmetric
scale (indicated), such that red luminescence reflects
poorly measured data). (b) Enlarged view of DNA
chromosomes.

analysis of DNA from cell lines containing
X chromosomes (Fig. 1b), as we did before
the sensitivity of our method to detect single-
(1- (47,XXX), 2- (48,XXXX), or
X) gains (also see Fig. 5, which is published
nation on the PNAS web site). Fluorescence
proportional to copy number ratios, which
estimated, in agreement with previous ob-
servations DNA copy number alterations were
breast cancer cell lines and primary tumors
in the tumors despite the presence of euploid
cells; the magnitudes of the observed changes
in the tumor samples. DNA copy-number
and in every cancer cell line and tumor, and
chromosome in at least one sample. Recurrent
copy number gain and loss were readily iden-
tifiable, gains within 1q, 8q, 17q, and 20q were
proportion of breast cancer cell lines/tumors
70%, 100%/60%, and 90%/44%, respective-
ly, within 1p, 3p, 8p, and 13q (80%/24%,
70%, and 70%/18%, respectively), consistent
genetic studies (refs. 2-4; a complete listing
provided in Tables 2 and 3, which are published
nation on the PNAS web site). The total

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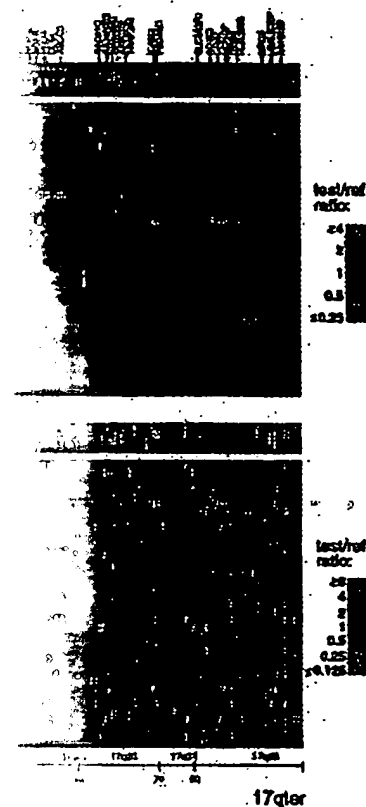


reillustrated for cell lines containing different numbers are separately ordered by hierarchical clustering to to chromosome 8 are ordered by position along the selected genes are indicated with color-coded text (red, red mRNA levels (observed in the majority of the subset not represented on the microarray are indicated in the for breast cancer cell line SKBR3. Fluorescence ratios

interval recurrently amplified in the tumors we case, known or plausible candidate oncogenes in description of these regions, as well as the red regions on chromosomes 17 and 20, can be and 7, which are published as supporting PNAS web site). breast cancer cell lines and tumors (4 and 37, a subset of arrayed genes (6,095), mRNA actively measured in parallel by using cDNA the parallel assessment of mRNA levels is pretation of DNA copy number changes. For ly amplified genes that are also highly ex- ingest candidate oncogenes within an ampli- significantly, our parallel analysis of DNA es and mRNA levels provides us the oppor- global impact of widespread DNA copy n gene expression in tumor cells. e of DNA copy number on gene expression mination of the pseudocolor representations

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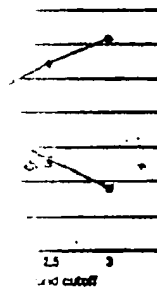
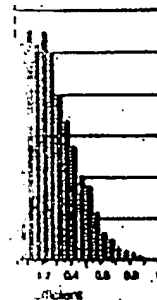
alteration (Upper) and mRNA levels (Lower)
by hierarchical clustering (Upper), and the
17, and for which both DNA copy number
in color-coded text (see Fig. 2 legend).

lines and tumors, average mRNA levels
copy number across all five classes, in a
fashion (P values for pair-wise Student's
cent classes: cell lines, 4×10^{-49} , 1×10^{-49} ,
tumors, 1×10^{-43} , 1×10^{-214} , 5×10^{-41} ,
regression of the average log(DNA copy
level, against average log(mRNA level)
average, a 2-fold change in DNA copy
died by 1.4- and 1.5-fold changes in mRNA
cell lines and tumors, respectively (Fig.
not shown). Second, we characterized the
25 correlations between DNA copy num-
each across the 37 tumor samples (Fig. 4b).
correlations forms a normal-shaped curve,
shifted in the positive direction from
tically significant, as evidenced in a plot
ed correlations (Fig. 4c), and reflects a
ate of DNA copy number alterations on
the highest correlations between DNA
level (the right tail of the distribution
with amplified and deleted genes (data not
a linear regression model to estimate the
measured in mRNA levels among the 37

Fig. 4. Mean and standard deviation of the log₂ ratio of tumor to normal expression for the 7% of genes that are up-regulated in tumor samples.

tumor copy number changes (7% of genes) are up-regulated in tumor samples. The mean and standard deviation of the log₂ ratio of tumor to normal expression for the 7% of genes that are up-regulated in tumor samples.

Fig. 4



(a) Distribution of correlations between observed versus expected correlation coefficients. The line of unity is indicated. (b) Percent variance retained (black line) and fraction of data retained (gray line) versus cutoff values. Fraction of data retained is relative to variation in gene expression attributable to copy number changes.

Although the DNA microarrays used in our study were biased toward characterized and/or highly expressed genes (we are examining such a large fraction of genes, approximately 20% of all human genes), and although we are likely underestimating the fraction of copy number changes to altered gene expression, our findings are likely to be generalizable. It will still be remarkable if only applicable to a subset of genes.

Gene copy number changes have been shown to result in gene expression biases (13). Two recent studies examine the global relationship between gene copy number and gene expression in cancer cells. In a study by Phillips *et al.* (14) have shown that chromosomal gains and losses in an immortalized prostate cell line result in significant respective increase and decrease in expression level of involved genes. In a study by Phillips *et al.* (15) recently reported that in metastatic prostate cancer, genes within amplified regions were 2-fold (or more) expressed, when compared with non-amplified genes. This report differs substantially from our findings, which show that highly amplified genes in breast cancer show decreased expression. These contrasting findings may be due to methodological differences between the

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of expressed genes, even within existing
on data sets, may permit the inference
erration, particularly aneuploidy (where
s averaged across large chromosomal
l supporting information). Fifth, this
substantial portion of the phenotypic
tension, the heterogeneity in clinical
ts' tumors may be traceable to underly-
py number. Sixth, this finding supports
pread DNA copy number alteration in
beyond the amplification of specific
of specific tumor suppressor genes.
number alteration, and the concomitant
gene expression, might disrupt critical
ips in cell metabolism and physiology
ic spindle), possibly promoting further
and directly contributing to tumor
sion. Finally, our findings suggest the
erapies that exploit specific or global
ession in cancer.

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Associate Investigator. This work was
e National Institutes of Health, the Howard
the Norwegian Cancer Society, and the
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